

# **Certificate of Analysis**

Product Cat. No.: M00593

**Host Cell:** CHO-K1

Target gene: BDCA2 and FcER1G

**Shipping Conditions:** Dry ice

**Lot Number:** B30191703



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### **Certificate of Analysis**

#### **Stable Cell Line Information**

Recommended Cell Culture Medium: F-12K (Gibco, Cat. #21127-022), FBS (Gibco, Cat. #10099-141), and

8µg/ml Puromycin\* (Gibco, Cat. #10131-027)

Freeze Medium: 95% complete growth medium, 5% (V/V) DMSO

Description: One stable subline using CHO-K1 as the host will be established to overexpress BDCA2 and

FcER1G

**QC:** FACS

Mycoplasma 160 Test\*\*: Negative

#### **Notice to Purchaser:**

GenScript stable cell line products are to be used for research purposes only, not intended for use in humans. GenScript products may not be transferred to third parties or used to manufacture commercial products without written approval. Use of this product is also subject to compliance with the licensing requirements.

<sup>\*</sup> Concentration used for selection was 8 µg/ml Puromycin.

<sup>\*\*</sup> Our PCR mycoplasma test covers 160 of the most common species of mycoplasma, with sufficient sensitivity and specificity.



# **Certificate of Analysis**

### **QC Data**

#### 1. Validation by Flow Cytometry

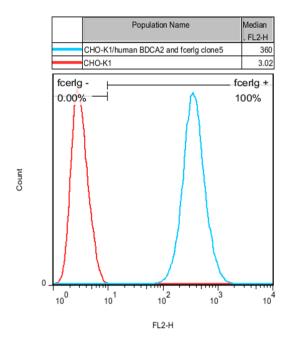


Figure 1. FACS analysis of BDCA2 and FcER1G in CHO-K1/BDCA2 and FcER1G

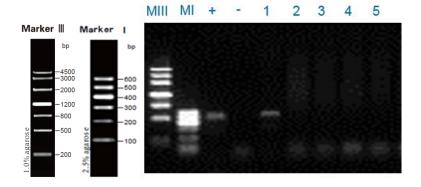


Figure 2. Myco 160 analysis of BDCA2 and FcER1G in CHO-K1/BDCA2 and FcER1G (Lane 05: M00593)



## Instruction for maintaining stable cell line

#### **Cell recovery**

The cells were maintained in F-12K, 10% FBS, and 8 µg/ml Puromycin. The S.O.P for cell recovery is briefly introduced here:

- 1) Prepare a 37°C water bath.
- 2) Take the cryovial out of the liquid nitrogen tank and thaw it by gentle swirling in the water bath until ice crystals are melted, usually within 1-3 minutes.
- 3) Decontaminate the vial by spraying with 70% ethanol.
- 4) Unscrew the vial and transfer the cells into a 15 ml sterile conical centrifuge tube containing 9 ml pre-warmed complete growth medium.
- 5) Centrifuge the tube at 125 g for 10 minutes, discard the supernatant and resuspend the cells in 2 ml complete growth medium. Pipette gently to loosen the pellet.
- 6) Transfer the cell suspension to the culture vessel with antibiotic free medium and mix well. Incubate the vessel at 37°C, 5% CO<sub>2</sub>.
- 7) Replace the medium with fresh culture medium the next day (with appropriate concentration of antibiotic).

### **Cell Maintenance and Subculturing**

Volumes listed below are for 10 cm dish, proportionally reduce or increase the volume for culture vessels of other sizes.

- 1) Balance the complete growth medium to 37°C in a water bath.
- 2) Discard the culture medium of the dish.
- 3) Briefly rinse the cell layer with  $Ca^{2+}/Mg^{2+}$  free DPBS to remove all traces of serum.
- 4) Add 1.0 to 2.0 ml of Trypsin-EDTA solution to the dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 1 to 3minutes).

Note: To avoid clumping, do not agitate the cells by hitting or shaking the dish while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate the dispersal.

- 5) Add 8.0 ml of complete growth medium and suspend the cells by gently pipetting.
- 6) Centrifuge the cells at 125 g for 10 minutes, discard the supernatant and resuspend the cells in 5 ml complete growth medium.



7) Add appropriate aliquot of the cell suspension to new culture vessel.

Subcultivation Ratio: 1:3 to 1:6

Medium Renewal: 2-3 times per week

### **Cell Cryopreservation**

1) Prepare a freeze medium consisting of complete growth medium and 5% DMSO.

- 2) Harvest cells by gentle centrifugation at 125 g for 10 minutes and resuspend them in the freeze medium at a concentration of  $1 \times 10^6$  to  $5 \times 10^6$  viable cells/ml. Rest of the cells are cultured until the viability of the recovered cells is confirmed.
- 3) Label the cryovials with the name of the cell line, then add 1 ml of the cell suspension to each of the vials.
- 4) Place the vials into a pre-cooled (4°C) controlled-rate freeze chamber and place the chamber in a -80°C freezer for at least 24 hours.
- 5) Quickly transfer the vials to a liquid nitrogen tank.
- 6) After 24 hours' preservation in liquid nitrogen, take one vial out and culture the cells to check the cell viability.



# Packing List of M00593

**Cell lines** (Shipping Condition: -80°C Dry Ice, Store at -196°C)

Name: CHO-K1/BDCA2 and FcER1G

**Quantity**: 1x10^6cells/vial

**Lot No.**: B30191703 **Number of vial**: 2 vials

Store at: -196°C

Certified by: Jan son

Date: <u>03/13/2018</u>

For research use only

860 Centennial Ave., Piscataway, NJ 08854, USA



# **Appendix**

1. Target gene information

BDCA2 and FcER1G, NM\_004106.1;

2. Antibodies used for FACS

Primary antibody: CD303(BDCA-2)-PE human (MACS,130-090-511)